REVIEW



Cepharanthine: a review of the antiviral potential of a Japanese-approved alopecia drug in COVID-19

Received: 12 May 2020 / Revised: 6 July 2020 / Accepted: 7 July 2020 / Published online: 22 July 2020 © The Author(s) 2020

Abstract

Cepharanthine (CEP) is a naturally occurring alkaloid derived from *Stephania cepharantha* Hayata and demonstrated to have unique anti-inflammatory, antioxidative, immunomodulating, antiparasitic, and antiviral properties. Its therapeutic potential as an antiviral agent has never been more important than in combating COVID-19 caused by severe acute respiratory syndrome coronavirus type 2 (SARS-CoV-2) virus. Cepharanthine suppresses nuclear factor-kappa B (NF-κB) activation, lipid peroxidation, nitric oxide (NO) production, cytokine production, and expression of cyclooxygenase; all of which are crucial to viral replication and inflammatory response. Against SARS-CoV-2 and homologous viruses, CEP predominantly inhibits viral entry and replication at low doses; and was recently identified as the most potent coronavirus inhibitor among 2406 clinically approved drug repurposing candidates in a preclinical model. This review critically analyzes and consolidates available evidence establishing CEP's potential therapeutic importance as a drug of choice in managing COVID-19 cases.

NIDO1

Keywords Cepharanthine · COVID-19 · Antiviral · Coronavirus · SARS-CoV-2

Abbreviations			
ACE2	Angiotensin converting enzyme 2		
CC_{50}	50% Cytotoxic concentration		
CEP	Cepharanthine		
CNS	Central nervous system		
COVID-19	2019 Novel coronavirus disease		
EC ₅₀	50% Effective concentration		
FDA	Food and drug administration		
HBV	Hepatitis B virus		
HCoV-OC43	Human coronavirus type OC43		
HIV	Human immunodeficiency virus		
HTLV	Human T-lymphocytic virus		
IC ₅₀	50% Inhibitory concentration		
IL-1β	Interleukin-1β		
IL-6	Interleukin-6		
LPS	Lipopolysaccharide		
NF-κB	Nuclear factor-kappa B		
NO	Nitric oxide		

Moshe Rogosnitzky moshe@medinsight.org

NPCI	Niemann-pick type C disease-causing	
	gene	
NSP	Non-structural protein	
SARS	Severe acute respiratory syndrome	
SARS-CoV	Severe acute respiratory syndrome	
	coronavirus	
SARS-CoV-2	Severe acute respiratory syndrome corona-	
	virus type 2 virus	
TMNAA	Tetrahydrotetramethylnaphthalene	
TNF-α	Tumor necrosis factor α	

Niamana mialatama C diasasa sansina

Introduction

The emergence of the 2019 novel coronavirus disease (labelled COVID-19) in December 2019 triggered an unprecedented global public health challenge. The disease is a clinical syndrome caused by the severe acute respiratory syndrome coronavirus type 2 (SARS-CoV-2) virus. With millions of cases globally, COVID-19 has become the world's largest and deadliest infectious disease outbreak since the Spanish flu of the early 1900s and it represents a serious public health problem that requires urgent solutions [1, 2]. So far, there is no confirmed cure for the disease which has led to hundreds of thousands of mortalities. The high transmission and exponential growth rate of the



MedInsight Research Institute, Pekeris 4, Weizmann Science Park, 7670204 Rehovot, Israel

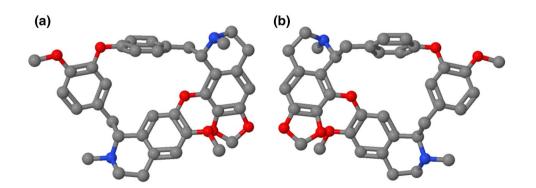
Institute for Personalized and Translational Medicine, Ariel University, 40700 Ariel, Israel

SARS-CoV-2 virus coupled with the slow process of developing vaccines and custom-designed pharmacological treatments have pointed at the need to quickly repurpose existing drugs such as cepharanthine (CEP) as potential treatments for COVID-19.

Cepharanthine is an alkaloid that has been used in Japanese medicine since 1951 primarily to treat radiationinduced leukopenia, alopecia areata, and alopecia pityrodes [3] as well as exudative middle-ear catarrh, and viper bite. It has been shown to possess anti-inflammatory, anti-oxidative, immuno-modulating, anti-parasitic, and antiviral properties which suggest possible use in a viral disease such as COVID-19 [3, 4]. More specifically, the antiviral effects of CEP have been demonstrated against HCoV-OC43, which is a mildly pathogenic human coronavirus [5], and severe acute respiratory syndrome coronavirus (SARS-CoV), which was responsible for the 2003 severe acute respiratory syndrome (SARS) outbreak [6]. Additionally, CEP was recently identified as the most effective drug against SARS-CoV-2-related pangolin coronavirus, a less pathogenic model for SARS-CoV-2, in a large drug screen of 2406 clinically approved drugs [7]. In the light of these findings and the close homology between the genome sequences of SARS-CoV and SARS-CoV-2 [2], CEP has become a drug of interest for treating COVID-19.

Extraordinary efforts are underway globally to identify safe and effective treatments for COVID-19. However, despite these efforts, only remdesivir currently has the United States Food and Drug Administration's (FDA) emergency use authorization for the treatment of COVID-19, and there is still limited information about the safety and effectiveness of this drug in clinical use [8, 9]. Chloroquine and hydroxychloroquine, which had earlier been granted FDA emergency use authorization in COVID-19, saw this authorization revoked due to lack of efficacy [10]. This further bolsters the need to critically examine and test the role of CEP as a potential anti-COVID-19 medication. This review is aimed at expediting this process by analyzing and consolidating published evidence on the antiviral effects of CEP and its therapeutic potential as a possible treatment for COVID-19.

Fig. 1 a, b Chemical structure of cepharanthine; front view (a) and back view (b)





A brief overview and current uses of cepharanthine

Cepharanthine is a natural alkaloid derived from plants of the genus Stephania native to Taiwan, China, Cambodia, Laos, Vietnam, and a few other southeastern Asian countries. Over the last 70–80 years, several members of the species have been used in Asia as traditional remedies for fever [4]. More specifically, *Stephania rotunda* Lour and *Stephania cepharantha* Hayata have been the most common species used as herbal medications in the region. There has been increasing interest in CEP because of its unique 1-benzylisoquinoline moiety (Fig. 1a, b), similarities with natural polypeptides, physiological properties, and long-established excellent safety profile [11].

Cepharanthine was first purified in 1934 by a Japanese pharmacist and named after one of its key sources: *Stephania cepharantha* [4]. Chemically, CEP is classified as a biscoclaurine alkaloid and is a member of the family of bisbenzylisoquinoline cyclic alkaloids which also includes tetandrine, daphnoline, berbamine, and several others [4]. In its chemical structure, CEP has two benzylisoquinoline units attached head-to-head which gives it an elliptic macrocyclic structure and confers it with its unique chemical properties such as its ether solubility, optical activity, and ability to reduce the fluidity of a wide variety of biological membranes [4, 12].

There have been several unsuccessful attempts at synthesizing CEP in a cost-effective manner [3]. So far, CEP has been optimally extracted using different patented approaches and formulated into orally administered tablets and powder, as well as injectables [4]. It is an approved medication indicated for managing radiation-induced leukopenia [13], alopecia areata [4], alopecia pityrodes [3], idiopathic thrombocytopenic purpura [14], middle ear catarrh, and bites from vipers and other venomous snakes [3]. At present, there is no clear consensus on the optimal dosing of CEP although doses of 2–60 mg per day have been used safely and effectively for the management of a wide variety of conditions [3]. Available data show that CEP has a half-life of 31.3–36.9 h with steady state levels

achieved after five to six repeated dosings of 100 mg/day and a maximum of 9.6% of unchanged drug is left following oral dosing [15]. The time to maximum serum concentration after oral administration of a single 10 mg to 60 mg dose in healthy adult males ranges between 1.1 and 2.5 h [3]. Following absorption, CEP is metabolized extensively in the liver and distributed to the tissues, where it binds strongly.

Broad pharmacologic effects of CEP have been demonstrated in vitro and in vivo using various experimental models. Ershun et al. demonstrated CEP's anti-inflammatory properties using an in vivo mouse model of mastitis and showing that CEP reduced the levels of tumor necrosis factor α (TNF- α), interleukin-1 β (IL-1 β), and interleukin-6 (IL-6) through an attenuation of the natural inflammatory response [16]. Similarly, an experiment using a mouse model of acute lung injury and lipopolysaccharide (LPS)-stimulated RAW264.7 cells showed that CEP does suppress nuclear factor-kappa B (NF- κ B) activation, lipid peroxidation, nitric oxide (NO) production, and expression of cyclooxygenase; thereby causing a dampened inflammatory response and subsequent inhibition of vascular smooth muscle cell proliferation and migration associated with atherosclerosis [17].

Also, CEP has been shown through both in vitro and in vivo experiments to prevent cell death by suppressing endotoxin-induced production of NO in macrophages; thereby protecting against widespread endotoxin damage in septic shock [18]. Other key effects of CEP include in vivo inhibition of superoxide generation and luminol-dependent

chemiluminescence in guinea-pig neutrophils [19], in vitro stimulation of autophagy and autophagic cell death in a panel of apoptosis-resistant cancer cells [20], in vivo and in vitro inhibition of vascular endothelial growth factor and IL-8 (which are key pro-angiogenic substances) and NF-κB activity [21]. Cepharanthine is also known to block *Plasmodium falciparum* development in its ring stages [22] and potentiate anti-cancer activity of chemotherapeutic agents by inhibiting drug transporters leading to the accumulation of the agents in cells [23].

Antiviral effects

Beyond the general pharmacological effects identified and discussed earlier, there have been in vitro studies pointing at specific antiviral properties of CEP that make it a potential drug of choice in the management of COVID-19 (Table 1).

In vitro effects of CEP on Human Immunodeficiency Virus (HIV)

Cepharanthine has been demonstrated to have unique antiviral properties on the human immunodeficiency virus (HIV). Okamoto et al. showed the extent to which CEP inhibits replication of HIV type 1 (HIV-1) in infected monocytic and lymphocytic cell lines [24]. At a 50% effective concentration (EC $_{50}$) of 0.016 µg/ml and a cytotoxic concentration (CC $_{50}$) of 2.2 µg/ml, the researchers found CEP to dose-dependently inhibit the replication of the virus in TNF- α -stimulated U1

Table 1 Summary of studies examining antiviral effects of cepharanthine

Virus	Cell type	Antiviral effect	Mechanism of action
HIV-1 [24]	U1 monocytes and ACH-2 T lymphocytes	Inhibits viral replication in U1 monocytic cells, but not in ACH-2 lymphocytic cells	Inhibits NF-κB activation
HTLV-1 [28]	MT-2 cells ^a SIT cells ^b	Inhibits the proliferation of both cell lines; and induces apoptosis of SIT cells	Inhibits NF-κB activation
HBV [30]	HepG2 cells ^c	Inhibits viral replication and suppresses viral HBeAg antigen production	Suppresses HBV via downregulation of host Hsc70 ^d expression
SARS-CoV [6]	VeroE6 ^e	Inhibits viral replication	Mechanism unknown
HCoV-OC43 [5]	MRC-5 fibroblasts	Inhibits viral replication and infectivity; and dampens virus-induced host response	Blocks expression of the viral spike protein and nucleoprotein
SARS-CoV-2 [1]	VeroE6/TMPRSS2	Inhibits viral entry and replication	Binds to the spike protein and interferes with viral engagement to ACE2
SARS-CoV-2 [34]	Vero cells	Suppresses viral infectivity	Mechanism unknown
SARS-CoV-2 [35]	Calu-3 human lung cells	Inhibits viral infectivity	Mechanism unknown
GX_P2V ^f [7]	VeroE6 ^e	Inhibits viral replication and cytopathic effects	Mechanism unknown

^aMT-2 is an HTLV-1-transformed T-cell line



^bSIT is a leukemia cell line established from patients with adult T-cell leukemia

^cHuman hepatoma cell line—HepG2

^dHeat stress cognate-70 is a host protein critical to HBV replication

^eMonkey kidney epithelial cell line VeroE6

^fPangolin coronavirus GX_P2V/pangolin/2017/Guangxi

1512 M. Rogosnitzky et al.

monocytic cells. This effect was attributed to CEP's ability to suppress HIV-1 long terminal repeat-driven gene expression by inhibiting the activation of NF- κ B (Fig. 2). Of all the cellular factors involved in HIV-1 replication and expression, NF- κ B is the most potent activator of its expression [25], and its inhibition by CEP goes a long way in dampening the virus's pathogenicity.

Additional in vitro experiments have gone further to compare CEP's anti-HIV-1 abilities with other alkaloid derivatives of cepharanoline. According to Baba et al. CEP was found to be the most active inhibitor of NF-kB (among 96 derivatives of cepharanoline) and thus, the most potent agent against HIV-1 activity in chronically infected cells [26]. The ability of CEP to easily penetrate the central nervous system (CNS) makes it even more useful in managing HIV-1-associated conditions of the CNS [27]. Compared with conventional antiviral agents currently used in the management of HIV (such as reverse transcriptase and protease inhibitors), CEP appears to be better suited to serve as both a

prophylactic and therapeutic agent in preventing and treating HIV-associated CNS disorders, primarily because it easily traverses the blood–brain barrier and can suppress the production of inflammatory cytokines that are responsible for CNS damage [24].

In vitro effects of CEP on human T-lymphocytic virus (HTLV)

Similar to the effects of CEP on HIV-1, the human T-lymphotropic virus type 1 (HTLV-1), which is the causative agent of adult T-cell leukemia, has been found to be susceptible to CEP. In vitro experiments by Toyama et al. show that the use of CEP alone or in combination with tetrahydrotetramethylnaphthalene (TMNAA) derivative triggered apoptosis of HTLV-1 infected cells through the caspase-dependent pathway [28]. The authors postulated that this effect may be due to the inhibition of the NF-κB signalling pathway found to be upregulated in infected cells (Fig. 2). The combination

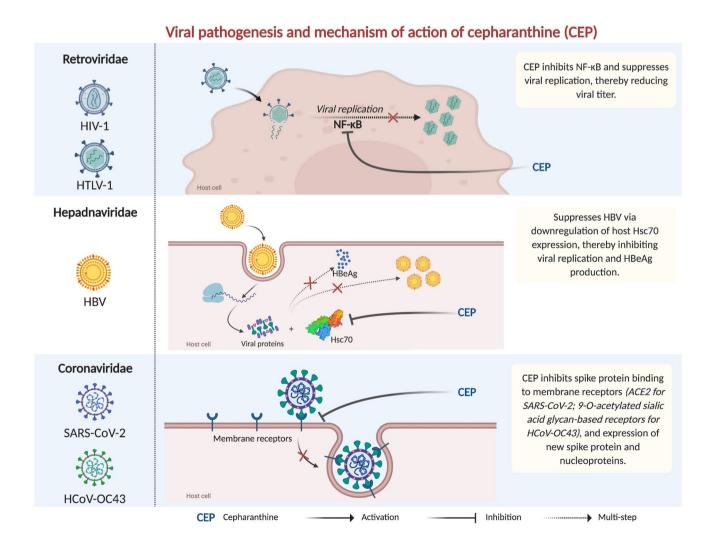


Fig. 2 Pathogenesis of HIV-1, HTLV-1, HBV, SARS-CoV-2, and HCOV-OC43; and mechanism of action of cepharanthine against them



of CEP with TMNAA in this study was observed to generate synergistic antiproliferative activity which suggests that the dose of either agent can be reduced to achieve the same or even better efficacy than the use of one agent alone. Furthermore, there is evidence pointing at the ability of CEP to reduce or reverse the resistance of cancer cells to antitumor agents, suggesting that combination chemotherapy with CEP has superior benefits [5, 28].

In vitro effects of CEP on hepatitis B virus (HBV)

Lamivudine is a nucleoside analogue commonly used as an antiviral agent against HBV. Over time, resistance against lamivudine has been on the rise with as much as 76% of patients developing drug resistance after 5 years of lamivudine therapy [29]. The need for alternatives has never been more apparent and CEP has proven useful in addressing this challenge. In a study comparing the antiviral effects of lamivudine, CEP, and adefovir on virus antigen production and HBV replication; lamivudine was found to have the most efficacy of all three substances when tested on wild-type HBV [30]. This was followed by CEP which had a higher rate of inhibition of both virus replication and viral antigen production than adefovir on both wild-type and lamivudine-resistant HBV subtypes. When tested on HepG2 cells, both CEP and adefovir exhibited similar cytotoxic effects.

In vitro effects of CEP on SARS-CoV and HCoV-OC43

Severe acute respiratory syndrome is a highly contagious atypical respiratory illness caused by the human coronavirus: SARS-CoV. Responsible for over 8000 deaths between 2002 and 2003, scientists have been working hard to find a cure [31]. Cepharanthine is an agent that has shown preclinical promise in the treatment of SARS. Zhang et al. conducted in vitro experiments to assess the extent of anti-SARS-CoV activity using VeroE6 cells infected with SARS-CoV [6]. The researchers split the cells into four groups, where cells in the first group were pre-treated with CEP prior to infection and cells in the second group were treated with CEP post-infection with the virus. The third group had cells cotreated with CEP and the virus. The fourth group of cells were also co-treated with CEP and the virus but the mixture of CEP and virus were incubated at 37 °C for 2 h prior to application on the VeroE6 cells. Following treatment, viral cytopathic effect was inhibited by a CEP concentration of 10 μg/ml across all groups, with the 50% inhibitory concentration (IC₅₀) ranging between 6.0 and 9.5 μ g/ml for the four treatments [6].

Even though Zhang et al. [6] had no clear explanation for their findings, their results were not totally surprising as CEP has also been demonstrated to inhibit similar human coronaviruses such as the human coronavirus type OC43

(HCoV-OC43). A 2019 study examined the effects of bisbenzylisoquinoline alkaloids (including CEP) on human MRC-5 lung cells infected with HCoV-OC43 [5]. Following pretreatment with CEP and HCoV-OC43, MRC-5 cells that were exposed to CEP prior to the viral inoculation showed no virus-induced cytotoxicity or morphological changes with the CC₅₀ and EC₅₀ found to be 10.54 μmol/L and 729.7 nmol/L, respectively. Another key finding from the study was the increase in the viability of infected cells in a dose-dependent manner [5]. These findings were attributed to the ability of CEP to inhibit viral RNA replication, block expression of viral proteins, and suppress production of proinflammatory molecules towards preventing an exacerbated cytokine response to the viral infection.

These findings are crucial to understanding the potential of CEP as an efficacious treatment option for COVID-19 particularly because of the similarities between HCoV-OC43, SARS-CoV, and SARS-CoV-2; all of which are human coronaviruses [32]. As pointed out previously, SARS-CoV-2 shares 79.6% gene sequence homology with SARS-CoV and both viruses utilize the same cell-membrane receptor—angiotensin converting enzyme 2 (ACE2)—to achieve viral entry [2] (Fig. 2). Unlike SARS-CoV and SARS-CoV-2, however, HCoV-OC43 uses 9-*O*-acetylated sialic acid glycan-based receptors to mediate viral entry [5]. Nonetheless, given the remarkable effect of CEP on viral binding, entry, and replication in the case of both HCoV-OC43 and SARS-CoV; there is a probability that CEP will show similar effects on the newly discovered SARS-CoV-2.

Effects of CEP on SARS-CoV-2 (in vitro and predictive models)

Recent studies indicate that CEP could be an effective treatment against SARS-CoV-2 with minimal toxicity. A group of researchers recently conducted a panel screening of drugs (including nelfinavir, CEP, and chloroquine) already approved for the treatment of various conditions, on an in vitro SARS-CoV-2 cell culture model [1]. Using VeroE6/ TMPRSS2 cells inoculated with SARS-CoV-2, nelfinavir and CEP were found to have excellent antiviral effects on SARS-CoV-2 either singly or in combination. Nelfinavir is a known protease inhibitor and the modeling data from the study confirmed that its ability to inhibit SARS-CoV-2 replication was due to inhibition of the virus' main protease. CEP, on the other hand, was found to predominantly block viral entry by interfering with the ability of the virus to attach to its target cell via its S glycoprotein [33]. When combined, both agents work synergistically to target both the viral entry and replication phases of SARS-CoV-2 infection, suggesting superior outcomes when compared with chloroquine and lopinavir which are among the currently used drugs for COVID-19 treatment [1]. Additionally, the EC_{50} and CC_{50}



ratio in this combination was over 70 (IC $_{50}$ 0.35 µmol/L, CC $_{50}$ 25.10 µmol/L) implying that SARS-CoV-2 inhibition can be achieved with minimal toxicity.

In yet another drug screening to repurpose already approved drugs into possible COVID-19 treatments, Fan et al. tested over 2400 drugs on VeroE6 cells infected with a 2019-nCoV-related pangolin coronavirus (GX_P2V/ pangolin/2017/Guangxi) as an alternative to SARS-CoV-2 because of its low human pathogenicity [7]. The spike protein of the pangolin coronavirus has 92.2% amino acid sequence similarity with the spike protein of the clinical isolate of SARS-CoV-2, thereby supporting its use as a model for SARS-CoV-2. Of the 2406 drugs tested; CEP, selamectin, and mefloquine hydrochloride were the only three drugs observed to completely inhibit the virus' cytopathic effect. Of these three drugs, CEP had the most potent antiviral activity even at low IC₅₀ (0.73 µmol/L) and CC₅₀ (39.30 μmol/L); similar to what had been reported in tests against SARS-CoV and HCoV-OC43 [5, 6]. Also, CEP was found to significantly decrease viral RNA yields across the entry and post-entry phases by as much as 2.17 fold and 1618 folds, respectively, relative to normal infections without any treatment [7]. This implies that CEP plays inhibitory roles both at the entry and post-entry phases of SARS-CoV-2 infection.

The antiviral potential of CEP against SARS-CoV-2 has been further demonstrated in recent in vitro studies using SARS-CoV-2 models. Jeon et al. who screened 3000 approved drugs for anti-SARS-CoV-2 activity, shortlisted CEP as one of 24 drugs that demonstrated antiviral activity against SARS-CoV-2 in Vero cells [34]. In a follow-up study by the same group utilizing Calu-3 cells, CEP ranked in fifth place with an IC $_{50}$ of 30 μ M (as opposed to an IC $_{50}$ of 4.47 μ M in Vero cells) [35].

Ruan et al. indicated that the antiviral activity of CEP against SARS-CoV-2 is due to its ability to combine well with a multi-subunit complex of nonstructural proteins (NSP) [36]. This conclusion was based on CEP's ability to combine well with the NSP12-NSP7-NSP8 complex in a virtual screening based on homologous models of SARS NSPs. Being an RNA virus, the NSP12-NSP7-NSP8 complex in SARS-CoV-2 is deemed critical for viral replication and transcription [37]. By binding to this NSP machinery, CEP blocks the ability of SARS-CoV-2 to replicate and produce relevant proteins. The inhibition of the Niemann-Pick type C disease-causing gene (NPC1) is another mechanism through which CEP has been shown to impair the infectivity of SARS-CoV-2 and subsequent replication [38]. As a NPC1 inhibitor, CEP causes disruption of cellular/lysosomal lipid homeostasis and potentially inhibits the entry, replication, and exit of SARS-CoV-2 [38, 39].

So far, encouraging preclinical evidence about CEP's safety profile, its availability, and unique antiviral properties

especially against specific beta-coronaviruses supports its consideration as a drug of choice in the treatment of human coronavirus diseases. In vivo testing and clinical trials are urgently required to provide additional data supporting the development of CEP into a COVID-19 treatment, spurred by its superior antiviral and antiinflammatory activities in preclinical settings relative to established antiviral agents. The ability of CEP to disable the attachment and entry of SARS-CoV-2 into target cells demonstrates its great therapeutic potential as a drug to combat COVID-19. Also, its low EC₅₀ and CC₅₀ achievable with oral dosing, and synergistic properties when combined with other antiviral medications (such as nelfinavir and mefloquine), further supports its excellent pharmacokinetic profile and establishes its importance as a candidate in the fight against COVID-19.

Conclusion

In summary, CEP has been demonstrated pre-clinically to have significant antiviral effects against viruses such as HIV, HTLV, HBV, SARS-CoV, and now, SARS-CoV-2, which is the culprit behind the world's largest and deadliest infectious disease outbreak within the last 100 years. Monotherapy with CEP has been demonstrated to inhibit viral entry and attenuate the potential inflammatory effects that may result from viral infection. These effects can be accentuated through combinations of CEP with other antiviral agents such as nelfinavir, mefloquine, or selamectin. Considering the fact that the majority of available literature on the antiviral effects of CEP have been from in vitro studies, further research on in vivo models and clinical trials are urgently called for to confirm the usefulness of CEP in real-life COVID-19 cases and enable the full maximization of the potentials of this naturally-occurring alkaloid. Given the well-established safety profile and clear antiviral and antiinflammatory effects of CEP, it presents as an optimal candidate for rapid repurposing to combat viral infection from SARS-CoV-2 and its COVID-19 sequelae.

Author contributions All authors contributed to the design of the work, literature reviews, manuscript development, and final approval of the version to be published.

Funding This research did not receive any sponsorship from funding agencies in public, commercial, or not-for-profit sectors. The Uniting History Foundation, Riga, Latvia, provided funding for Open Choice costs.



Compliance with ethical standards

Conflict of interest The authors declare no conflicts of interest. Moshe Rogosnitzky is a named inventor on patents utilizing cepharanthine for treatment of androgenic alopecia.

Open Access This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit http://creativecommons.org/licenses/by/4.0/.

References

- Ohashi H, Watashi K, Saso W, Shionoya K, Iwanami S, Hirokawa T, et al. Multidrug treatment with nelfinavir and cepharanthine against COVID-19. BioRxiv. 2020. https://doi. org/10.1101/2020.04.14.039925.
- Zhou P, Yang XL, Wang XG, Hu B, Zhang L, Zhang W, et al. A
 pneumonia outbreak associated with a new coronavirus of probable bat origin. Nature. 2020;579(7798):270–3.
- Rogosnitzky M, Danks R. Therapeutic potential of the biscoclaurine alkaloid, cepharanthine, for a range of clinical conditions. Pharmacol Rep. 2011;63:337–47.
- Bailly C. Cepharanthine: an update of its mode of action, pharmacological properties and medical applications. Phytomedicine. 2019;62:152956.
- Kim DE, Min JS, Jang MS, Lee JY, Shin YS, Park CM, et al. Natural bis-benzylisoquinoline alkaloids-tetrandrine, fangchinoline, and cepharanthine, inhibit human coronavirus OC43 infection of MRC-5 human lung cells. Biomolecules. 2019;9(11):696.
- Zhang CH, Wang YF, Liu XJ, Lu JH, Qian CW, Wan ZY, et al. Antiviral activity of cepharanthine against severe acute respiratory syndrome coronavirus in vitro. Chin Med J. 2005;118(6):493–6.
- Fan HH, Wang LQ, Liu WL, An XP, Liu ZD, He XQ, et al. Repurposing of clinically approved drugs for treatment of coronavirus disease 2019 in a 2019-novel coronavirus (2019-nCoV) related coronavirus model. Chin Med J. 2020;133(9):1051–6.
- Food and Drug Administration. Coronavirus (COVID-19) Update: FDA issues emergency use authorization for potential COVID-19 treatment. FDA News Release [Internet]. 2020. https://www.fda. gov/news-events/press-announcements/coronavirus-covid-19update-fda-issues-emergency-use-authorization-potential-covid -19-treatment. Accessed 21 June 2020.
- Sanders JM, Monogue ML, Jodlowski TZ, Cutrell JB. Pharmacologic treatments for coronavirus disease 2019 (COVID-19): a review. JAMA. 2020. https://doi.org/10.1001/jama.2020.6019.
- Food and Drug Administration. Coronavirus (COVID-19) update: FDA revokes emergency use authorization for chloroquine and hydroxychloroquine [Internet]. 2020. https://www.fda.gov/newsevents/press-announcements/coronavirus-covid-19-update-fdarevokes-emergency-use-authorization-chloroquine-and. Accessed 21 June 2020.
- Kulka M. Bisbenzylisoquinoline alkaloids. In: Manske R, editor. The alkaloids chemistry and physiology. London: Academic Press; 1965. p. 233.

- Nagatsuka S, Nakazawa T. Effects of membrane-stabilizing agents, cholesterol and cepharanthine, on radiation-induced lipid peroxidation and permeability in liposomes. Biochim Biophys Acta (BBA)-Biomembr. 1982;691(1):171–7.
- Nomoto S, Imada H, Ohguri T, Yahara K, Kato F, Morioka T, et al. Effect of Cepharanthine in preventing radiation induced normal tissue damage in prostate cancer. Gan To Kagaku Ryoho. 2004;31(7):1063–6.
- Nakayama S, Matsushita A, Ichiba S, Nagai K. Clinical evaluation of cepharanthin for chronic idiopathic thrombocytopenic purpura. Rinsho Ketsueki. 1992;33(3):408–9.
- Yasuda K, Moro M, Akasu M, Ohnishi A. Pharmacokinetics of Cepharanthin in Phase I clinical trials (single and continuous intravenous administration). Jpn J Clin Pharmacol Ther. 1989;20(4):741-9.
- Ershun Z, Yunhe F, Zhengkai W, Yongguo C, Naisheng Z, Zhengtao Y. Cepharanthine attenuates lipopolysaccharide-induced mice mastitis by suppressing the NF-κB signaling pathway. Inflammation. 2014;37(2):331–7.
- 17. Paudel KR, Karki R, Kim D-W. Cepharanthine inhibits in vitro VSMC proliferation and migration and vascular inflammatory responses mediated by RAW2647. Toxicol In Vitro. 2016;34:16–25.
- Sakaguchi S, Furusawa S, Wu J, Nagata K. Preventive effects of a biscoclaurine alkaloid, cepharanthine, on endotoxin or tumor necrosis factor-alpha-induced septic shock symptoms: involvement of from cell death in L929 cells and nitric oxide production in raw 264.7 cells. Int Immunopharmacol. 2007;7:191–7.
- Tsuyoshi M, Kunzoorita O, Keisuke E, Hirotsugu K, Eisuke FS, Bunji I, et al. Inhibition of active oxygen generation in guineapig neutrophils by biscoclaurine alkaloids. Biochem Pharmacol. 1990;39(7):1255–9.
- Law BYK, Chan WK, Xu SW, Wang JR, Bai LP, Liu L, et al. Natural small-molecule enhancers of autophagy induce autophagic cell death in apoptosis-defective cells. Sci Rep. 2014;4:5510.
- Harada K, Ferdous T, Itashiki Y, Takii M, Mano T, Mori Y, et al. Cepharanthine inhibits angiogenesis and tumorigenicity of human oral squamous cell carcinoma cells by suppressing expression of vascular endothelial growth factor and interleukin-8. Int J Oncol. 2009;35(5):1025–35.
- Desgrouas C, Dormoi J, Chapus C, Ollivier E, Parzy D, Taudon N. In vitro and in vivo combination of cepharanthine with antimalarial drugs. Malar J. 2014;13:90.
- Zhou Y. Cepharanthine is a potent reversal agent for MRP7(ABCC10)-mediated multidrug resistance. Biochem Pharmacol. 2009;77:993–1001.
- Okamoto M, Ono M, Baba M. Potent inhibition of HIV type 1 replication by an antiinflammatory alkaloid, cepharanthine, in chronically infected monocytic cells. AIDS Res Hum Retroviruses. 1998;14:1239–45.
- 25. Baba M. Cellular factors as alternative targets for inhibition of HIV-1. Antiviral Res. 1997;33(3):141–52.
- Baba M, Okamoto M, Kashiwaba N, Ono M. Anti-HIV-1 activity and structure-activity relationship of cepharanoline derivatives in chronically infected cells. Antivir Chem Chemother. 2001;12:307–12.
- Yokoshima T, Tsutsumi S, Ohtsuki T, Takaichi M, Nakajima T, Akasu M. Studies on metabolic fate of cepharanthine absorption, distribution, metabolism and excretion in rats. Iyakuhin Kenkyu. 1986;17:458–79.
- Toyama M, Hamasaki T, Uto T, Aoyama H, Okamoto M, Hashmoto Y, et al. Synergistic inhibition of HTLV-1-infected cell proliferation by combination of cepharanthine and a tetramethylnaphthalene derivative. Anticancer Res. 2012;32(7):2639–45.



1516 M. Rogosnitzky et al.

Yuen M-F, Fung J, Wong DK-H, Lai C-L. Prevention and management of drug resistance for antihepatitis B treatment. Lancet Infect Dis. 2009;9(4):256–64.

- 30. Zhou YB, Wang YF, Zhang Y, Zheng LY, Yang XA, Wang N, et al. In vitro activity of cepharanthine hydrochloride against clinical wild-type and lamivudine-resistant hepatitis B virus isolates. Eur J Pharmacol. 2012;683(1–3):10–5.
- Petrosillo N, Viceconte G, Ergonul O, Ippolito G, Petersen E. COVID-19, SARS and MERS: are they closely related? Clin Microbiol Infect. 2020;26(6):729–34. https://doi.org/10.1016/j. cmi.2020.03.026.
- St-Jean JR, Jacomy H, Desforges M, Vabret A, Freymuth F, Talbot PJ. Human respiratory coronavirus OC43: genetic stability and neuroinvasion. J Virol. 2004;78(16):8824

 –34.
- Lan J, Ge J, Yu J, Shan S, Zhou H, Fan S, et al. Structure of the SARS-CoV-2 spike receptor-binding domain bound to the ACE2 receptor. Nature. 2020;581(7807):215–20. https://doi. org/10.1038/s41586-020-2180-5.
- 34. Jeon S, Ko M, Lee J, et al. Identification of antiviral drug candidates against SARS-CoV-2 from FDA-approved drugs. Antimicrob Agents Chemother. 2020;64(7):e00819–20.
- Ko M, Jeon S, Ryu WS, Kim S. Comparative analysis of antiviral efficacy of FDA-approved drugs against SARS-CoV-2 in human lung cells: Nafamostat is the most potent antiviral drug candidate. bioRxiv [Internet]. 2020. https://www.biorxiv.org/

- content/10.1101/2020.05.12.090035v1.abstract. Accessed 21 June 2020.
- Ruan Z, Liu C, Guo Y, et al. SARS-CoV-2 and SARS-CoV: virtual screening of potential inhibitors targeting RNA-dependent RNA polymerase activity (NSP12). J Med Virol. 2020. https://doi.org/10.1002/jmv.26222.
- Yin W, Mao C, Luan X, Shen D-D, Shen Q, Su H, et al. Structural basis for inhibition of the RNA-dependent RNA polymerase from SARS-CoV-2 by remdesivir. Science. 2020;368(6498):1499–504. https://doi.org/10.1126/science.abc1560.
- Sturley SL, Rajakumar T, Hammond N, Higaki K, Márka Z, Márka S, Munkacsi AB. Potential COVID-19 therapeutics from a rare disease: weaponizing lipid dysregulation to combat viral infectivity. J Lipid Res. 2020;61(7):972–82. https://doi. org/10.1194/jlr.R120000851.
- Lyu J, Yang EJ, Head SA, Ai N, Zhang B, Wu C, et al. Pharmacological blockade of cholesterol trafficking by cepharanthine in endothelial cells suppresses angiogenesis and tumor growth. Cancer Lett. 2017;28(409):91–103.

Publisher's Note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

